

Improving Disease Diagnostics and Management Strategies of Recurrent Pathogens in California Greenhouses and Nurseries

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Outline

- How to scout for diseases in a nursery
- Update on improvement of diagnostics tools for Phytophthora detection

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- How to scout for diseases in a nursery
- Update on improvement of diagnostics tools for Phytophthora detection

Monitor



Patterns

Look for unusual trends

Are there symptoms?

How are they distributed ?



Symptom distribution



Same symptoms observed along the same area (bench)

Photo credit: Derby canyon Native Nursery

<https://derbycanyonnatives.com/2021/a-too-hot-summer-in-the-nursery-welcome-the-fall/>



Symptom distribution: biotic vs. abiotic

- Patches



- Complete areas,
whole crop



Symptoms caused by biotic factors

- Foliar symptoms can be the consequence of below ground rots



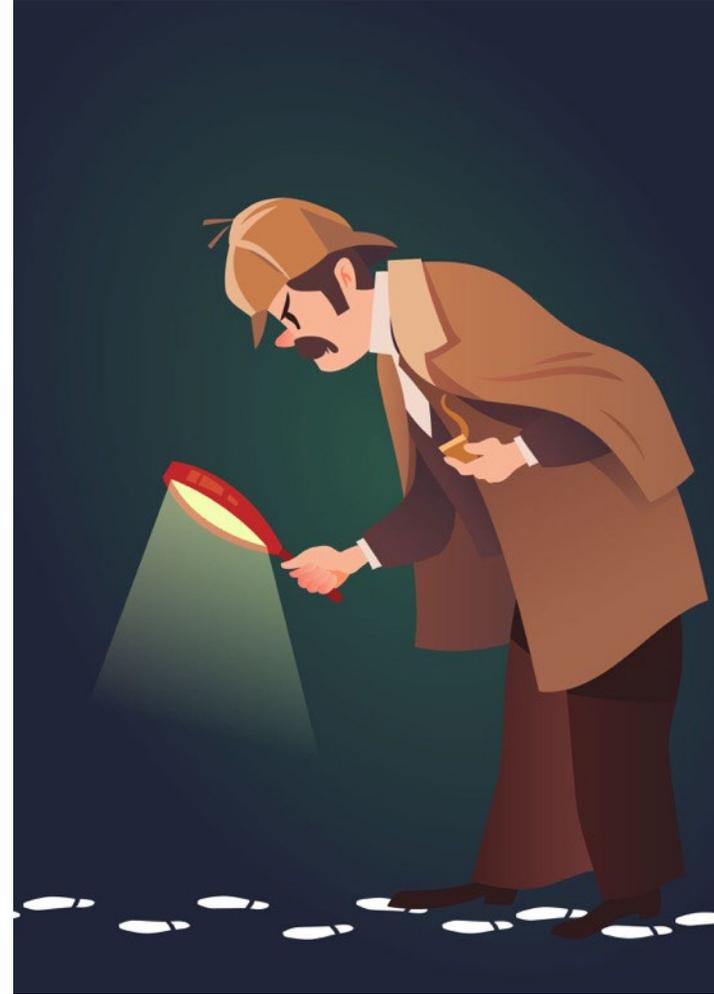
Environmental and cultural conditions

- Host: Crop age, cultivar
- Season → Temperature, RH
- Cultural practices
 - Irrigation
 - Standing water
 - Direct contact with soil
 - Plant spacing



Gathering “clues”

- Symptom distribution
- Symptoms observed
- Host:
 - Crop age
 - Cultivar
- Environment: temp., RH
- Cultural practices



1. Contact area farm advisor
2. Send diseased samples to a diagnostics lab!

Sample information

Plant _____ Variety _____ Date planted _____

Sample came from: Field/Farm Orchard
 Nursery Greenhouse Vegetable garden
 Christmas tree farm Other:

Soil characteristics: sandy clay loam
 organic hard pan soilless media
 other:

Symptoms: Root: Rot Lesions
Crown/stem/canopy: Rot Lesions Die back Wilt
 Vascular discoloration Canopy bleaching
Leaves: Speckled/ spots Marginal necrosis Mottle
 Chlorosis Deformed Mildew
Flowers: Rot Lesions Color break Deformed
Other symptoms _____

Exposure: full sun partial shade shade
 full shade windy protected

Irrigation type and frequency: _____

Chemicals applied: _____

Date damage first noticed: _____ **Number of years at present site:** _____

% Affected: _____ **Acreage:** _____

Number of plants affected (for potted plants) _____

Distribution of affected plants: single plant grouped scattered edge of field

Previous crops: _____

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How to submit samples?

- Fresh plant material
(not entirely dead plants!)
- When possible send the entire plant
- Wrap samples in paper towels
Do NOT add moisture
- Mail samples in a sturdy container:
Boxes, not bags
- Include pictures of the disease stages



(images credit: NDSU Plant diagnostic lab)

What is NOT a good diagnostics sample?



- Dead plants

What is NOT a good diagnostics sample?



- Parts of the plant instead of the whole plant
- Soil

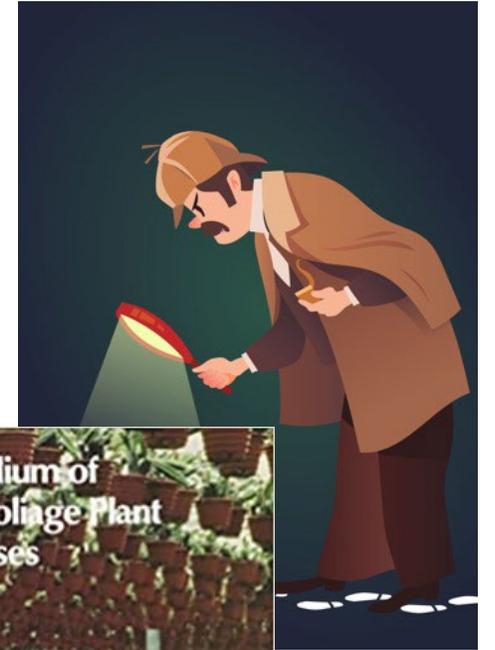
What is a good diagnostics sample?



- PLANTS CANNOT BE DEAD
- More than one plant (n= 3) plants
 - Foliar symptoms critical
- Capture range of symptoms
 - Minor, moderate, severe

Pathogen diagnostics

- Hypothesis based diagnostics:
 - Gathering clues: symptoms, host, environmental and cultural conditions
 - Disease compendiums, literature
- Tentative diagnostics (Hypothesis)
- Pathogen detection



Case study: Phytophthora detection

- Phytophthora spp. recovered from 20 of 26 surveyed nurseries:
 - *P. tentaculata* (27%)
 - *P. cactorum* (22%)
 - *P. cryptogea*-complex (13%)
- Hosts:
 - Toyon (26%)
 - Coffeeberry (17%)
 - Pacific madrone (8%)
 - California wild rose (8%)



Phytophthora in native habitats working group

An Accreditation Program to Produce Native Plant Nursery Stock Free of *Phytophthora* for Use in Habitat Restoration

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Phytophthora species appear to have wide host ranges, capable of causing disease on plants



- Implement Best Management Practices (BMPs) to produce native plant nursery stock free of *Phytophthora* for use in habitat restoration
 - Systematic approach to disease prevention that is integrated with nursery production practices

Preventing hazards in the nursery

1. Propagation source : Clean seed collection or shoot tip cuttings
2. Contaminated ground : Use of raised (75 cm) benches and gravel
3. Contaminated irrigation water: Use of well or city water
4. Used containers : Steam sterilization of containers
5. Contaminated potted media: Steam sterilization of media



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Protecting Native Habitats by Excluding Phytophthora in the Nursery

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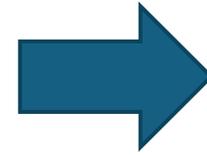
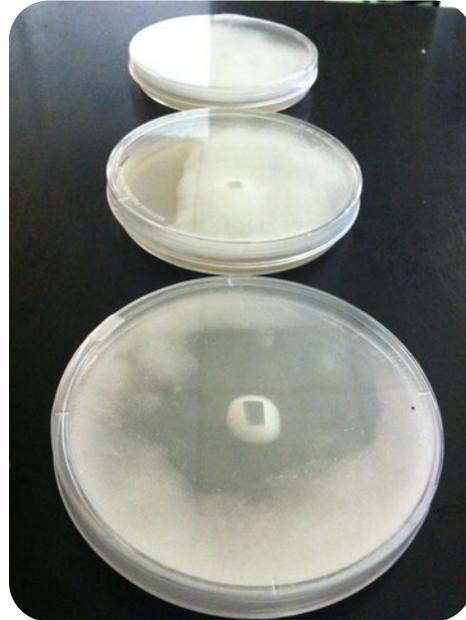
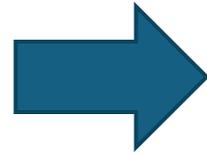
AIR Team Presents at the UC Davis

What is the most effective method for Phytophthora detection?

- Leachate method (sampling water)
- Sample from roots (SM isolation)
- Immunostrips



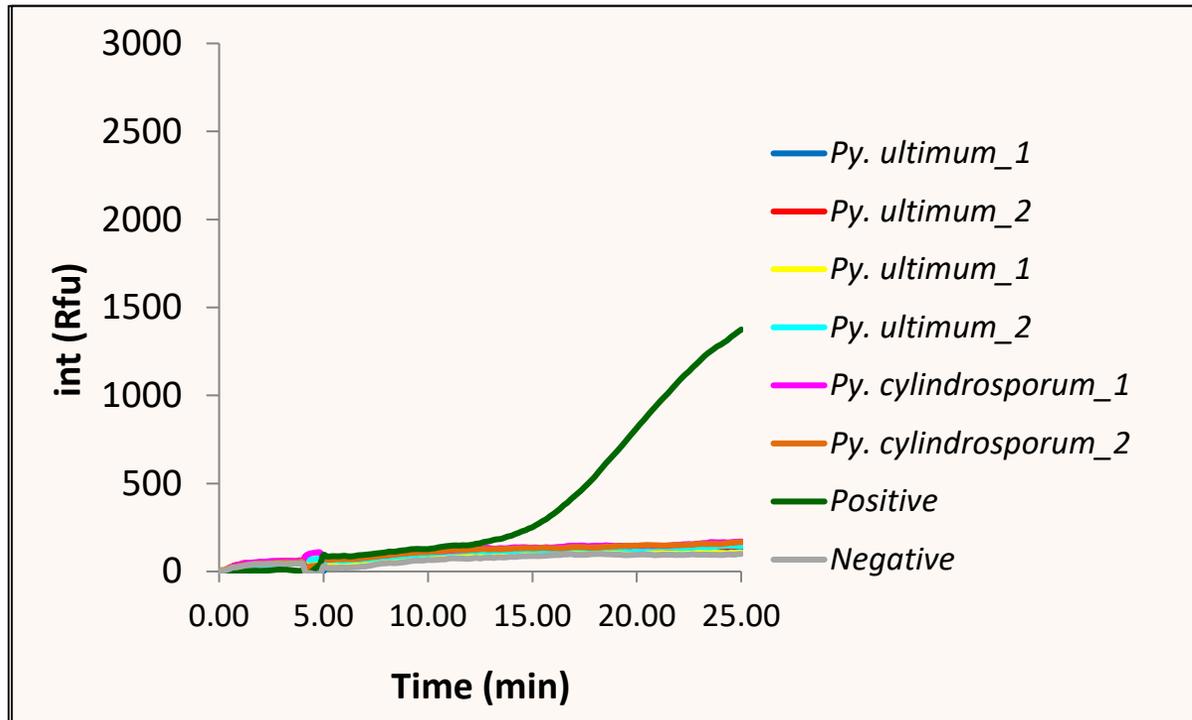
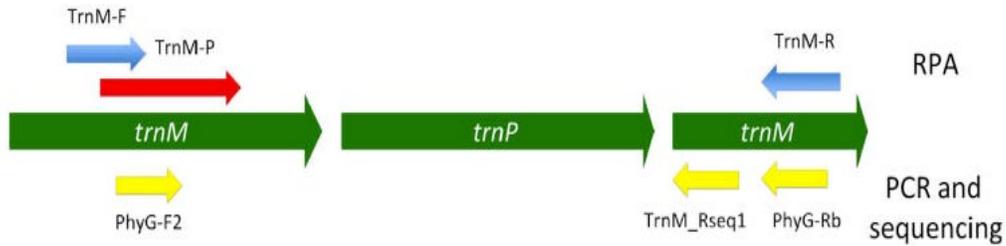
Phytophthora identification



Days for sample processing and final diagnostics

Are there rapid and more robust tools to detect *Phytophthora* ?

Recombinase Polymerase Amplification (RPA)



Dr. Frank Martin and Dr. Tim Miles

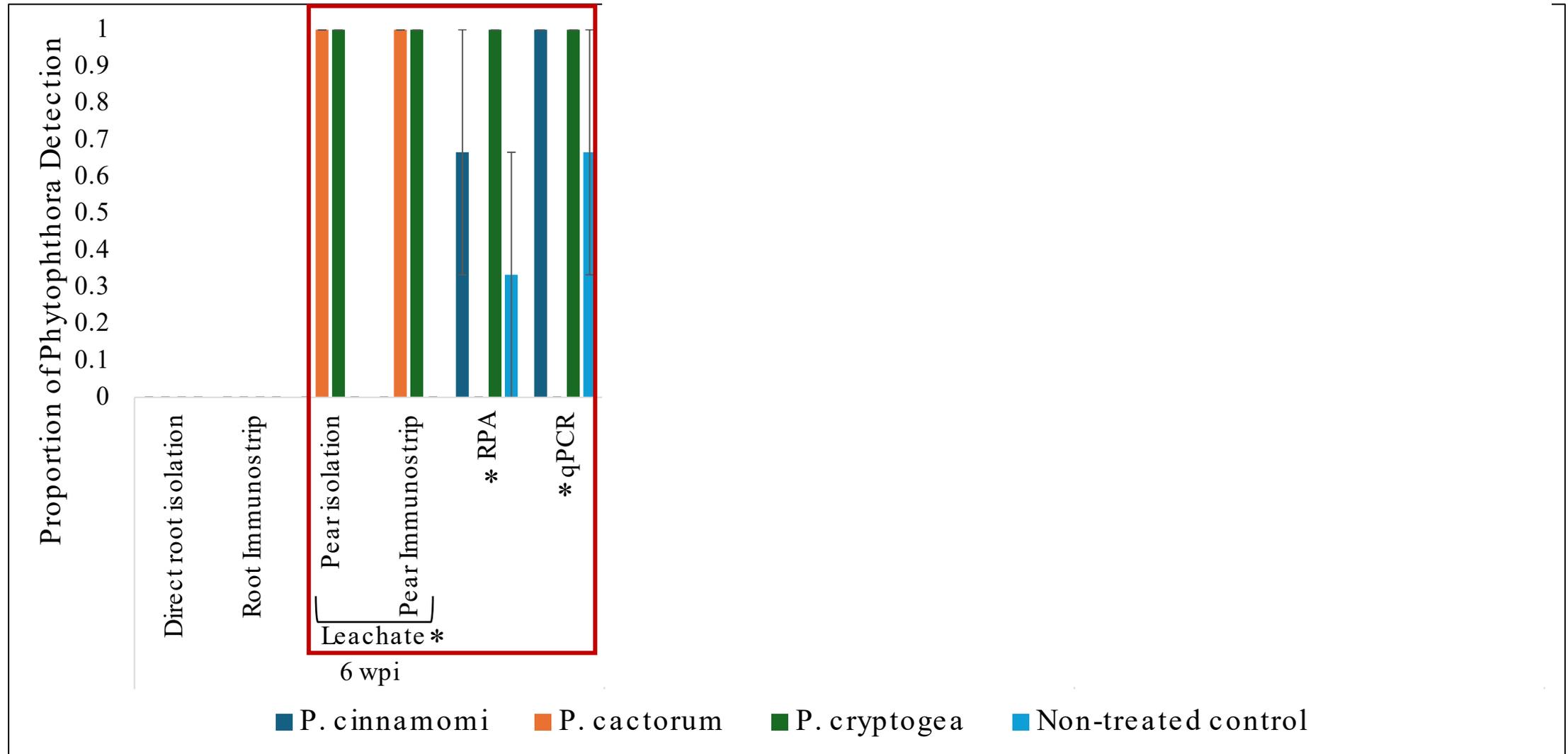
What is the most effective method for Phytophthora detection?



Randomized complete block design experiment

- 2 Plants
 - Coffeeberry and Toyon
- 4 Pathogen treatments
 - *P. cinnamomi*, *P. cactorum*, *P. cryptogea*, NTC
- 3 Sampling time points
 - 6 weeks post inoculation (wpi), 12 wpi. and 18 wpi.

Proportion of Phytophthora detected in inoculated coffeeberry using different detection methods (P <0.0001)



What is the most effective method for Phytophthora detection?

RPA, qPCR, and leachate baiting assay (pear isolations and pear immunostrips) had significantly greater detection rate than direct root isolation and root Immunostrips

How will the systems work for natural plant infestations?

Symptomatic plants sampled from 3 commercial and 2 native plant nurseries across California

On-site irrigation leachate testing was conducted in native plant nurseries



Arctostaphylos
'Howard McMinn'



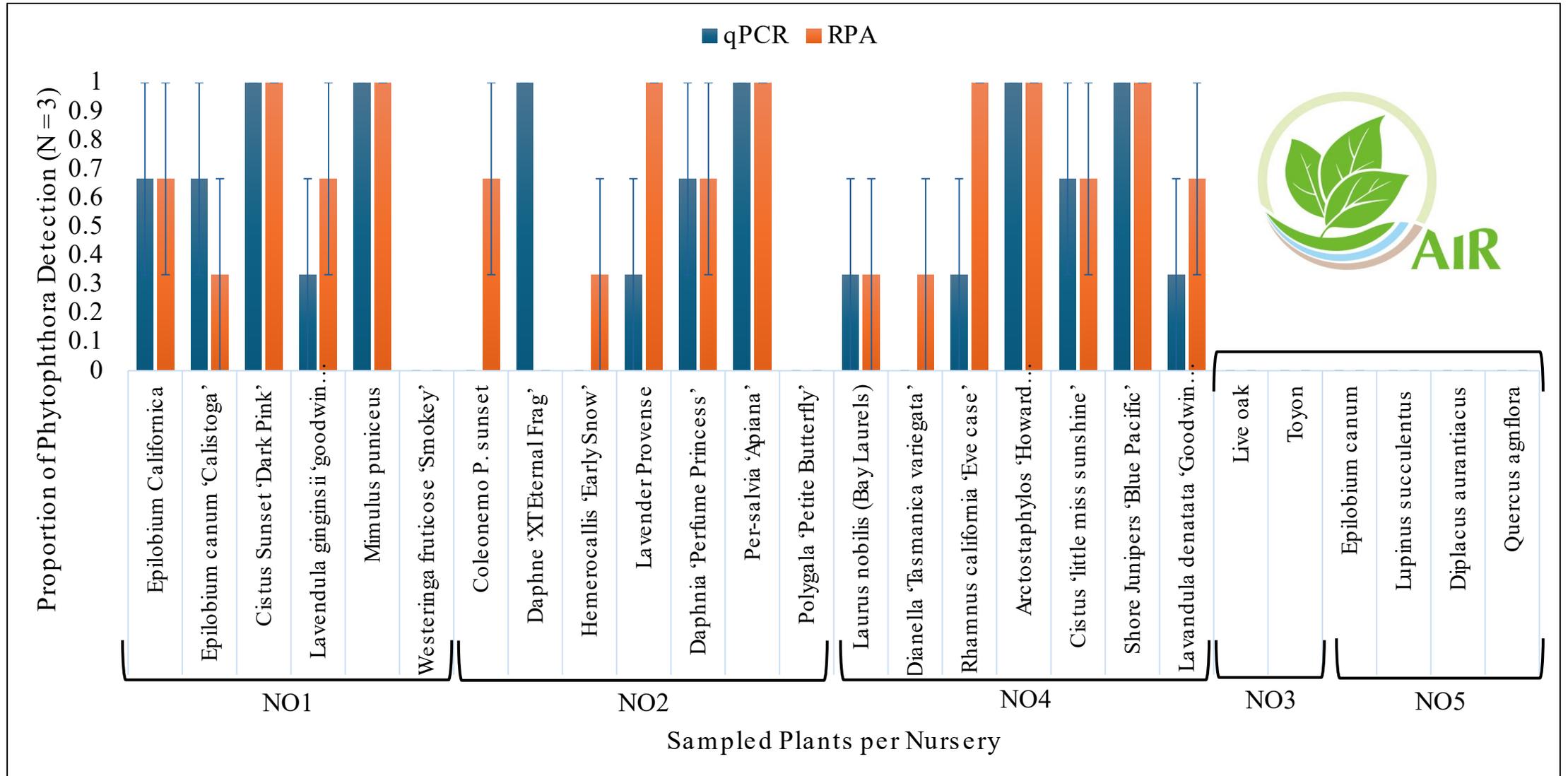
Cistus 'Little Miss
Sunshine'



Diplacus
aurantiacus



Proportion of *Phytophthora* detected in naturally infested plants showing crown and root rot symptoms sampled from five different nurseries, using RPA and qPCR assay methods



Conclusions

- RPA, qPCR, and irrigation leachate pear baiting had significantly higher *Phytophthora* spp. detection irrespective of host, pathogen species, and sampling time
- The RPA assay could be a robust on-site *Phytophthora* detection tool, and qPCR could be a robust lab-based assay for confirmatory detection of *Phytophthora*

Strict Best Management Practices along with timely diagnosis can help prevent *Phytophthora* spread into native ecosystems and landscapes

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Thank you!
Questions ?



<https://greenhousepathology.faculty.ucdavis.edu/>

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